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2'-Deoxyisoinosine: Synthesis of a Highly Fluorescent Nucleoside and Its Incorporation into Oligonucleotides

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2'-DEOXYISOINOSINE: SYNTHESIS OF A HIGHLY FLUORESCENT NUCLEOSIDE AND ITS INCORPORATION INTO OLIGONUCLEOTIDES

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ABSTRACT: The synthesis of 2'-deoxyisoinosine (2a) and the related 2',3'-dideoxynucleosides 2b and 3 is reported. The 3'-phosphonate 4b as well as the phosphoramidite 4c were prepared and employed in solid-phase oligonucleotide synthesis.

The 2'-deoxyinosine (1) is used as ambiguous nucleoside in oligonucleotide chemistry. We have synthesized the isomeric 2'-deoxyisoinosine (2a)¹ as well as the sugar derivatives 2b and 3. As an intermediate the aminonucleoside 6 was prepared. It was obtained from 2'-deoxyguanosine (4) via the 6-thioxo compound 5 by desulfurization². Treatment of 6 with NaNO₂/AcOH yielded 2a.

The synthesis of 2',3'-didehydro-2',3'-dideoxyisoinosine (3) was performed starting from 2'-deoxyisoinosine (2a). This was silylated to give compound 7.

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Mesylation afforded 8 containing two mesyl groups, one at the OH-3' group and the other at the oxygen-2. This was proved by ¹³C-NMR spectra of **2a** and **8** which exhibit a completely altered pattern of ¹³C-NMR resonances of the base. Moreover, compound **8** is no longer fluorescent. Compound **8** was then treated with Bu₄NF to give **3**. Subsequently, it was hydrogenated to 2',3'-dideoxyisoinosine (**2b**).

The 2-oxo nucleosides 2a, b and 3 show strong fluorescence (excitation: 315 nm, emission: 380 nm, in H_2O and MeOH). Treatment of the compounds 2a, c^3 with xanthine oxidase furnished the corresponding xanthine derivatives. The related 7-deaza-2'-deoxyisoinosine (2d) was found to be resistant towards the enzyme.

As it is unknown which of the common nucleosides show base pairing with 2a,c, oligonucleotides were synthesized. For this purpose, the nucleoside 2a was protected at OH-5' with a DMT-residue (9) and subsequently converted into its 3'-phosphonate 10a (a: $PCl_3 / 1,2,4$ -triazole; 31P-NMR: 0.52 J(P, H) = 587 Hz) as well as into the phosphoramidite 10b (b: Cl-P((i-Pr)₂N)O(CH₂)₂CN; 31P-NMR: 149.0, 148.3).

The polymer-linked 2'-deoxyisoinosine 10d was also prepared from 9 by succinylation (10c) followed by condensation with amino-functionalized CPG. The ligand concentration was 90 μ mol/g. ¹³C-NMR data are shown in the Table.

Table.	¹³ C-NMR	Data of 2	'-deoxyisoinosine	and derivatives	in d ₂ -DMSO.
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Compd.	C-2	C-6 139.4 142.7 ^t 139.1 139.5 139.5 139.3	C-5 123.6 123.2 123.5 123.6 123.6 123.6	C-8 145.5 142.5 ^t 145.3 145.7 145.4 145.5	C-4 158.8 159.5 158.7 159.0 158.9 159.0	CH ₃ O/CH ₃	
2a 3 7 9 10a 10c	156.1 157.5 155.9 155.9 156.0 155.8					25.9/18. 55.1 55.1 55.1	1
Compd.	C-1'	C-2'	C-3'	C-4'	C-5'	СООН	СО
2a 3 7 9 10a 10c	82.9 87.9 82.5 82.4 82.7 82.8	125.6	70.9 134.4 70.2 70.5 72.5 74.8	87.9 87.1 87.1 85.8 80.5 83.6	61.7 62.9 63.2 64.2 63.9 65.0	173.5	171.5

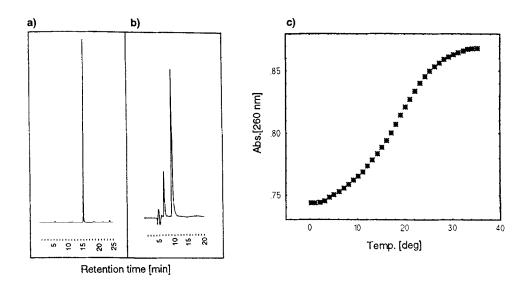


FIG. a) HPLC profile of 11, gradient 0-20% MeCN in 0.1 M (Et₃NH)OAc (pH 7.0) / MeCN, 95:5, at 260 nm; b) HPLC profile of the enzymatic digest of 11, at 290 nm; c) Melting curve of $11\cdot12$, at 260 nm.

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The phosphonate 10a was employed in solid-phase synthesis of the oligomers 11-13 using the phosphonate chemistry. HPLC profiles are shown in the Figure.

Next, the duplex formation of the oligonucleotide 11 with compounds 12 and 13 was studied. In both cases cooperative melting profiles were observed (Figure). A T_m value of 19°C was found for 11·12 and 20°C for 11·13. The related $d(A)_{12} \cdot d(T)_{12}$ showed a much higher T_m (44°C). This indicates that 2'-deoxyisoinosine does not base pair, neither with 2'-deoxythymidine nor with 2'-deoxyguanosine.

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